Pfam: multiple sequence alignments and HMM-profiles of protein domains

Erik L. L. Sonnhammer*, Sean R. Eddy¹, Ewan Birney², Alex Bateman² and Richard Durbin²

Computational Biology Branch, National Center for Biotechnology Information, National Library of Medicine, Building 38A, Room 8N805, National Institutes of Health, Bethesda, MD 20894, USA, ¹Department of Genetics, Washington University School of Medicine, St Louis, MO 63110, USA and ²Sanger Centre, Hinxton Hall, Cambridge CB10 1SA, UK

Received September 8, 1997; Revised and Accepted October 8, 1997

ABSTRACT

Pfam contains multiple alignments and hidden Markov model based profiles (HMM-profiles) of complete protein domains. The definition of domain boundaries, family members and alignment is done semi-automatically based on expert knowledge, sequence similarity, other protein family databases and the ability of HMM-profiles to correctly identify and align the members. Release 2.0 of Pfam contains 527 manually verified families which are available for browsing and on-line searching via the World Wide Web in the UK at http://www.sanger.ac.uk/ Pfam/ and in the US at http://genome.wustl.edu/Pfam/ Pfam 2.0 matches one or more domains in 50% of Swissprot-34 sequences, and 25% of a large sample of predicted proteins from the *Caenorhabditis elegans* genome.

INTRODUCTION

A relatively small number of structural and functional domains are used in a large number of different proteins. Particularly for protein analysis and annotation in large-scale sequencing projects, there is a growing need for easily interpretable and sensitive detection of common protein domains. A protein containing one or more common domains can produce a morass of hundreds or thousands of BLAST hits when searching single sequence databases (e.g. GenBank, Swissprot, PIR). Although searches can be augmented by tools that condense and summarise results (1), satisfactory annotation of such proteins often becomes a timeconsuming and error-prone process. Instead, a search of an organised database of protein domain families can produce more concise results which simplify annotation, domain parsing and functional prediction for a query sequence (2-5). Protein family databases are typically based on multiple sequence alignments of known family members. Conserved features can be recognised in the alignment and given higher weight in searches, which for distant similarities can often render the comparison more sensitive than pairwise alignment approaches.

We present here Pfam (6) release 2.0. Pfam was developed in order to use HMM-profile analysis to complement BLAST analysis in the *Caenorhabditis elegans* genome project. The main distinction between Pfam and most other protein family databases is that for all of Pfam, both the family definition and the search method span entire domains, including not only conserved motifs but also

less-conserved regions, insertions and deletions. HMM-profile methods allow variable conservation and insertions/deletions to be dealt with in a fairly robust way (7,8). Modelling of complete domains should facilitate more biologically meaningful sequence annotation, and, in some cases, more sensitive detection.

DESCRIPTION OF THE DATA

For each protein domain family in Pfam, there are three important files. The *seed alignment* is a manually verified multiple alignment of a representative set of sequences (Fig. 1). An *HMM-profile* is built from the seed alignment for database searching and alignment purposes. A *full alignment* is generated automatically from the seed HMM-profile by searching Swissprot for all detectable members and aligning them to the HMM-profile. The distinction between seed and full alignments facilitates updating the database; the seed alignments are stable resources, whereas full alignments and HMM-profiles can be generated automatically for any new Swissprot (or other sequence database) release.

Each family has a name, a permanent accession number and a record of the methods used to identify the family members and create the alignments. There is also either a brief description of the usual function and structure of the domain, or (more often) links to other on-line documentation resources such as Prosite and Prints.

Both the seed and the full alignments are subjected to a small array of 'quality control' procedures, to verify that the alignments are sensible, that the HMM-detected sequences in the full alignment include all presumed members of the family in Swissprot and no other sequences, and that the family does not overlap with other Pfam families. The process of generating the Pfam family is iterated, if necessary, until all quality requirements are met.

Most Pfam families are based on, and cross-referenced to, corresponding Prosite or Prints entries. In many cases, however, the definition of which sequences belong to a family differs between the databases. This is a pragmatic consequence of the different search methods used. Prosite and Prints detection relies primarily on short conserved patterns corresponding to superfamily motifs. A Prosite pattern or Prints fingerprint may recognise a highly conserved motif shared amongst an otherwise highly diverged superfamily that Pfam splits into several families; conversely, Pfam may recognise a superfamily that Prosite and Prints classify into several distinct families with distinct motif signatures. For some protein domain families, there may be no motif sufficiently conserved to make a

*To whom correspondence should be addressed. Tel: +1 301 435 5930; Fax: +1 301 480 9241; Email: sonnhammer@ncbi.nlm.nih.gov

ID	SH2								
AC	PF00017								
DE	Src homology de	lomain 2							
AU	Sonnhammer ELL	·							
AL	Clustalw								
AM	hmma -qR								
SE	Swissprot_feat	ure_table							
GA	Bic_raw 25 hmmfs 20								
DR	PROSITE; PDOC50001;								
DR	SCOP; 1sha; sf	· · · · · · · · · · · · · · · · · · ·							
SQ	58								
ABL1	_CAEEL/179-254	WYHGKISRSDSEAILGSGITGSFLVRESETSIGQYTISVRHDGRVFHYRINVDNTEK	MFITQEVKFRTLGELVHH						
BLK_	MOUSE/117-198	WFFRTISRKDAERQLLAPMNKAGSFLIRESESNKGAFSLSVKDITTQGEVVKHYKIRSLDNGG	YYISPRITFPTLQALVQHY						
BTK_	HUMAN/281-362	WYSKHMTRSQAEQLLKQE.GKEGGFIVRDS.SKAGKYTVSVFAKSTGDPQGVIRHYVVCSTPQS.Q	YYLAEKHLFSTIPELINYE						
CSW_	DROME/111-186	WFHGNLSGKEAEKLILERGK.NGSFLVRESQSKPGDFVLSVRTDDKVTHVMIRWQDKK	YDVGGGESFGTLSELIDHY						
CSW	DROME/6-81	WFHPTISGIEAEKLLQEQGF.DGSFLARLSSSNPGAFTLSVRRGNEVTHIKIQNNGDF	FDLYGGEKFATLPELVQY						
CTK	HUMAN/122-196	WFHGKISGQEAVQQLQPPEDGLFLVRESARHPGDYVLCVSFGRDVIHYRVLHRDGH	LTIDEAVFFCNLMDMVEHY						
DRK	DROME/60-134	WYYGRITRADAEKLLSNKHEGAFLIRISESSPGDFSLSVKCPDGVQHFKVLRDAQSK	.FFLWVVKFNSLNELVEY						
FER	HUMAN/460-531	WYHGAIFRIEAQELLKKQGDFLVRESHGKPGEYVLSVYSDGQRRHFIIQYVDNM	YRFEG.TGFSNIPQLIDH						
FPS_	DROME/438-510	WFHGVLPREEVVRLLNNDGDFLVRETIRNEESQIVLSVCWNGH.KHFIVQTTGEGN	FRFEG.PPFASIQELIMH						
FPS_	FUJSV/511-581	WYHGAIPRSEVQELLKYSGDFLVRESQGKQEYVLSVLWDGQPRHFIIQAADNL	YRLED.DGLPTIPLLIDH						
FRK	HUMAN/116-193	WFFGAIGRSDAEKQLLYSENKTGSFLIRESESQKGEFSLSVLDGAVVKHYRIKRLDEGG	FFLTRRRIFSTLNEFVSHY						
GTPA	HUMAN/181-256	WYHGKLDRTIAEERLRQAGK.SGSYLIRESDRRPGSFVLSFLSQMNVVNHFRIIAMCGD	YYIGG.RRFSSLSDLIGY						
GTPA	HUMAN/351-426	WFHGKISKQEAYNLLMTVG.QVCSFLVRPSDNTPGDYSLYFRTNENIQRFKICPTPNN	QFMMGGRYYNSIGDIIDH Y						
NCK_	HUMAN/282-356	WYYGKVTRHQAEMALNER.GHEGDFLIRDSESSPNDFSVSLKAQGKNKHFKVQLKET	VYCIGORKFSTMEELVEHY						
P85A	_HUMAN/624-698	WNVGSSNRNKAENLLRGKRDGTFLVRES.SKQGCYACSVVVDGEVKHCVINKTATGY	GFAEPYNLYSSLKELVLHY						
P85B	_BOVIN/618-692	WYVGKINRTQAEEMLSGKRDGTFLIRES.SQRGCYACSVVVDGDTKHCVIYRTATGF	GFAEPYNLYGSLKELVLHY						
PIP4	RAT/668-741	WYHASLTRAQAEHMLMRVPR.DGAFLVRKR.NEPNSYAISFRAEGKIKHCRVQQEGQ	TVMLGNSEFDSLVDLISY						
SEM5	_CAEEL/60-136	WYLGKITRNDAEVLLKKPTVRDGHFLVRQCESSPGEFSISVRFQDSVQHFKVLRDQNGK	.YYLWAVKFNSLNELVAY						
SHC_	HUMAN/378-449	WFHGKLSRREAEALLQLNGDFLVRESTTTPGQYVLTGLQSGQPKHLLLVDPEG	VVRTKDHRFESVSHLISY						
SRC1	_DROME/162-244	WFFENVLRKEADKLLLAEENPRGTFLVRPSEHNPNGYSLSVKDWED.GRGYHVKHYRIKPLDNGG	YYIATNQTFPSLQALVMA						
SRC2	_DROME/214-292	WYVGYMSRQRAESLLKQG.DKEGCFVVRKS.STKGLYTLSLHTKVPQSHVKHYHIKQNARCE	YYLSEKHCCETIPDLINY						
SRK1	_SPOLA/122-199	WFLGKIKRVEAEKMLNQSFNQVGSFLIRDSETTPGDFSLSVKDQDRVRHYRVRRLEDGS	LFVTRRSTFQILHELVDHY						
SRK4	_SPOLA/122-199	WFFGQVKRVDAEKQLMMPFNNLGSFLIRDSDTTPGDFSLSVRDIDRVRHYRIKKLENGT	YFVTRRLTFQSIQELVAY						
STK_	HYDAT/126-203	WYFGDVKRAEAEKRLMVRGLPSGTFLIRKAETAVGNFSLSVRDGDSVKHYRVRKLDTGG	YFITTRAPFNSLYELVQH						
SYK_	HUMAN/15-92	FFFGNITREEAEDYLVQGGMSDGLYLLRQSRNYLGGFALSVAHGRKAHHYTIERELNGT	YAIAGGRTHASPADLCHY						
SYK_	PIG/163-238	WFHGKISRDESEQIVLIGSKTNGKFLIRARDNGSYALGLLHEGKVLHYRIDKDKTGK	LSIPGGKNFDTLWQLVEHY						
TEC_	MOUSE/246-329	WYCRNTNRSKAEQLLRTE.DKEGGFMVRDS.SQPGLYTVSLYTKFGGEGSSGFRHYHIKETATSPKK	YYLAEKHAFGSIPEIIEYH						
TXK_	HUMAN/150-231	WYHRNITRNQAEHLLRQE.SKEGAFIVRDS.RHLGSYTISVFMGARRSTEAAIKHYQIKKNDSGQ	WYVAERHAFQSIPELIWY						
VAV_	MOUSE/671-745	WYAGPMERAGAEGILTNRSDGTYLVRQRVKDTAEFAISIKYNVEVKHIKIMTSEGL	YRITEKKAFRGLLELVEF?						
YES_	KIPHE/159-241	WYFGKLSRKDTERLLLLPGNERGTFLIRESETTKGAYSLSLRDWDE.TKGDNCKHYKIRKLDNGG	YYITTRTQFMSLQMLVKHY						
YKF1	_CAEEL/20-101	YFHGLIQREDVFQLLDNNGDYVVRLSDPKPGEPRSYILSVMFNNKLDENSSVKHFVINSVENK	YFVNNNMSFNTIQQMLSH						
ZA70	_HUMAN/163-239	WYHSSLTREEAERKLYSGAQTDGKFLLRPRK.EQGTYALSLIYGKTVYHYLISQDKAGK	YCIPEGTKFDTLWQLVEY						
ZA70	MOUSE/10-87	FFYGSISRAEAEEHLKLAGMADGLFLLRQCLRSLGGYVLSLVHDVRFHHFPIERQLNGT	YAIAGGKAHCGPAELCOF						

Figure 1. Example of a typical Pfam entry, the SH2 family. Shown is the flat file record including a reduced version of the seed alignment.

discriminative pattern or fingerprint. (Prosite is increasingly incorporating profiles for these families; these Prosite profiles are very similar to Pfam models.) Only the largest (>15 members) Prosite families were systematically used to construct Pfam entries. For smaller families, constructing an HMM-profile is of less value since the sensitivity is unlikely to improve relative to singlesequence searching, and because a small sample is often nonrepresentative. Of the 71 Pfam families with no corresponding Prosite or Prints entry, 55 were 'discovered' as large clusters in Pfam-B (see below). 24 Pfam families contain links to other World Wide Web (WWW) protein family documentation resources, some of which were gleaned from the ProWeb server (9).

Pfam 2.0 contains 527 families, comprising 39 113 sequence segments and 6.8 million residues in the full alignments. All sequences were taken from Swissprot 34(10). The alignments are on average 275 residues wide, including gaps. There are on average ~75 members per family in full alignments, and ~22 in seed alignments.

Pfam-B

For comprehensiveness, all Swissprot sequences not in Pfam are clustered automatically by the program Domainer (2), which also constructs multiple alignments automatically and is the basis for the ProDom protein family database. The quality of these alignments tends to be low, but domain-based automated clustering is a convenient method of identifying large obvious families that need to be targeted for Pfam model construction. Although we do not stably maintain, annotate or produce HMM-profiles of these clusters, we make them available as Pfam-B. Pfam-B 2.0 contains 13 289 clusters, 62 611 subsequences, and 8.2 million residues. On average, alignments are 146 residues wide (including gaps) and contain five members.



Figure 2. Pfam 2.0 contains domains from nearly half of all Swissprot 34 proteins. The automatic clusters in Pfam-B 2.0 contain domains from 33% of the Swissprot proteins that do not contain Pfam domains. When counting residue-by-residue, roughly a third of Swissprot is covered by Pfam and Pfam-B each. Pfam-B does not include proteins known to be fragments or segments shorter than 30 residues; the figures for unique sequences are therefore overestimated.

Sequence database coverage

As shown in Figure 2, 48% of the sequences and 32% of the residues in Swissprot 34 are included in annotated Pfam alignments. If unannotated Pfam-B clusters are also taken into account, 81% of sequences and 71% of residues in Swissprot 34 are included in Pfam. In searches of a large and presumably unbiased set of predicted protein sequences from the *C.elegans* genome, 25% of sequences and 13% of residues show significant hits to Pfam HMM-profiles. The numbers are slightly lower for prokaryotic genomes.

SEARCHING Pfam

The US and UK Pfam WWW servers provide users the ability to search query protein sequences against one, all, or a few Pfam

b

a	Score	Query from	Query to	HMM from	HMM to	Pfam Family	Description
	97.67	104	153	1	50	DAG_PE-bind	Phorbol esters / diacylglycerol binding domain
	92.44	169	218	1	50	DAG_PE-bind	Phorbol esters / diacylglycerol binding domain
	137.88	240	328	1	92	C2	C2 domain
	276.16	413	674	1	247	pkinase	Eukaryotic protein kinase domain
	\$4.44	675	741	1	69	pkinase_C	Protein kinase C terminal domain
	70.99	807	857	17	69	pkinase_C	Protein kinase C terminal domain



Figure 3. Tabular output (a) and schematic output (b) from a Pfam search with the *C.elegans* protein E01H11.1 as query. Both pictures were taken from the Washington University WWW server.

HMMs. Results are returned in tabular format, and both GIF- and Java-based graphical representations are available optionally. An example of the results from such a search is shown in Figure 3. Here, the *C.elegans* Kin-11 gene product (E01H11.1) is shown to possess a duplicated phorbol esters/diacylglycerol binding domain (DAG/ PE-bind), a C2 domain, a protein kinase catalytic domain (pkinase) and a duplicated domain frequently associated C-terminally to protein kinase domains (pkinase_C).

Users can also use Pfam HMM-profiles to search protein sequences locally using the freely available HMMER software package at http://genome.wustl.edu/eddy/hmmer.html#hmmer For comparing genomic and EST data to Pfam HMM-profiles, the programs GeneWise and ESTWise (11) are available at http://www.sanger.ac.uk/Software/Wise2/

WORLD WIDE WEB SERVERS, FTP ACCESS AND FORMAT

The Pfam home pages are http://www.sanger.ac.uk/Pfam/ at the Sanger Centre in the UK and http://genome.wustl.edu/Pfam/ at Washington University in the USA. The two servers are separately maintained and differ slightly in their services and capabilities, but are based on the same underlying Pfam database. Both servers support HMM searching, browsing of the family alignments and documentation and lookup of the domain organisation of proteins in Swissprot.

The entire database, including accessory data files such as Pfam schematics for Swissprot proteins, is also available as flat file format ASCII files by anonymous FTP at ftp.sanger.ac.uk and genome.wustl.edu in /pub/databases/Pfam/

The format of the Pfam alignment flat files is based on the EMBL/Swissprot two-character field labels. The following Pfam-specific labels are used: AL, alignment method of seed members; AM, alignment method of full alignment; AU, author responsible for the alignments; GA, gathering method/search program and cutoffs used to build full alignment; SE, source suggesting the seed members belong to the same family; SQ, number of sequences (and last line before the alignment starts). The alignment is in a simple format (Fig. 1) which consists of one line per subsequence containing the Swissprot sequence ID, start and end of the segment, and the aligned subsequence itself (no length limit). In the Pfam flat file, the corresponding Swissprot accession number is added to the right of each alignment line. Users of the Pfam database or WWW servers should cite this article as the appropriate reference.

ACKNOWLEDGEMENTS

We thank Robert Finn for preparing most of the new families for Pfam 2.0, and Jose Aguilar for writing and maintaining the Washington University Pfam server. Pfam development in SRE's group is supported by grant R01-HG01363 from the NIH National Human Genome Research Institute. Pfam development at the Sanger Centre is supported by the Wellcome Trust.

REFERENCES

- Sonnhammer, E.L.L. and Durbin, R. (1994) Comput. Appl. Biosci., 10, 301–307
- 2 Sonnhammer, E.L.L. and Kahn, D. (1994) Protein Sci., 3, 482-492.
- 3 Attwood, T.K., Beck, M.E., Bleasby, A.J., Degtyarenko, K., Michie, A.D. and Parry-Smith, D.J. (1997) *Nucleic Acids Res.*, 25, 212–217 [see also this issue (1998) *Nucleic Acids Res.* 26, 304–308].
- 4 Bairoch, A., Bucher, P. and Hofmann, K. (1997) Nucleic Acids Res., 25, 217–221.
- 5 Henikoff,J.G., Pietrokovski,S. and Henikoff,S. (1997) Nucleic Acids Res., 25, 222–226 [see also this issue (1998) Nucleic Acids Res. 26, 309–312].
- 6 Sonnhammer, E.L.L., Eddy, S.R. and Durbin, R. (1997) *Proteins*, **28**, 405–420.
- 7 Krogh,A., Brown,M., Mian,I.S., Sjoelander,K. and Haussler,D. (1994) J. Mol. Biol., 235, 1501–1531.
- 8 Eddy, S.R. (1996) Curr. Opin. Struct. Biol., 6, 361-365.
- 9 Henikoff,S., Endow,S.A. and Greene,E.A. (1996) Trends Biochem. Sci., 21, 444–445.
- 10 Bairoch, A. and Apweiler, R. (1997) Nucleic Acids Res., 25, 31–36 [see also this issue (1998) Nucleic Acids Res. 26, 38–42].
- 11 Birney, E. and Durbin, R. (1997) In ISMB-97; Proceedings Fifth International Conference on Intelligent Systems for Molecular Biology. AAAI Press, Menlo Park, pp. 56–64.